

**NOVA SCOTIA AQUACULTURE REVIEW BOARD**

IN THE MATTER OF: *Fisheries and Coastal Resources Act, SNS 1996, c 25*

- and -

IN THE MATTER OF: An Application by KELLY COVE SALMON LTD. for a boundary amendment and two new finfish aquaculture licenses and leases for the cultivation of Atlantic salmon (*Salmo salar*) - AQ#1205x, AQ#1432, AQ#1433, in Liverpool Bay, Queens County (the "**Application**")

**Affidavit of Shawn Robinson, PhD affirmed on January 19, 2024**

I affirm and give evidence as follows:

1. I am Shawn Robinson, PhD of St. Andrews, New Brunswick. I was a research scientist with the Government of Canada, Department of Fisheries and Oceans until my retirement in 2022. I am currently a senior scientist with Longline Environment, a UK research and innovation company providing services to a variety of industries, including aquaculture.
2. I have personal knowledge of the evidence affirmed in this affidavit except where otherwise stated to be based on information and belief.
3. I state, in this affidavit, the source of any information that is not based on my own personal knowledge, and I state my belief of the source.
4. I have been retained by Kelly Cove Salmon Limited ("**KCS**") to provide my independent expert opinion to the Nova Scotia Aquaculture Review Board in connection with KCS's Application to expand its Atlantic salmon operations at Coffin Island (AQ#1205X) and for two new Atlantic salmon aquaculture farms at Mersey Point (AQ#1433) and Brooklyn Point (AQ#1432).
5. In particular, I have been asked for my independent expert opinion with respect to the effect of the KCS's proposed expansion of its salmon aquaculture operations on the American lobster population in Liverpool Bay.
6. My independent opinion is set out in my report attached as **Exhibit A**.
7. My CV is attached as **Exhibit B**.

**VIRTUALLY AFFIRMED** before me in Halifax, Nova Scotia, on MS Teams with Mr. Robinson in St. Andrews, New Brunswick on January 19, 2024.



---

Barrister of the Supreme Court of Nova Scotia



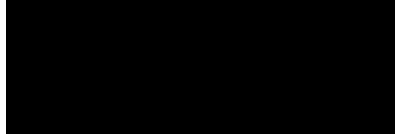
---

Shawn Robinson, PhD

# TAB A

**KCS' Application re AQ#1205X, AQ#1432,  
AQ#1433 in Liverpool Bay, Queens County**

This is Exhibit A referred to in the Affidavit  
of Shawn Robinson, PhD, affirmed virtually  
before me on January 19, 2024.



---

Barrister of the Supreme Court of Nova Scotia

# ARB Review Submission – Lobster/Farm Interactions

Shawn Robinson, Ph.D.  
Longline Environment, 63 St Mary Axe, London, EC3A 8AA, United Kingdom

## Introduction

1. I have been retained by Kelly Cove Salmon Limited (“KCS”) to provide my independent expert opinion to the Nova Scotia Aquaculture Review Board in connection with KCS’s Application to expand its Atlantic salmon operations at Coffin Island (AQ#1205X) and for two new Atlantic salmon aquaculture farms at Mersey Point (AQ#1433) and Brooklyn Point (AQ#1432).
2. In particular, I have been asked for my independent expert opinion with respect to the effect of the KCS’s proposed expansion of its Atlantic salmon aquaculture operations in Liverpool Bay on the American lobster population in Liverpool Bay.

## Historical Background

3. Canada has been a dominant player in the marine food production industry and has a long history of fishing dating back over 500 years with the activity of the early Portuguese fishers off the Grand Banks of Newfoundland<sup>1</sup>. To understand where we are going with marine food production, it is valuable to have some insight on where we have been. In the 1950’s, Canada ranked 5<sup>th</sup> in the world for total fisheries production, but over time, that rank has diminished to the point where Canada is now 28<sup>h</sup> in the world as of 2021 and

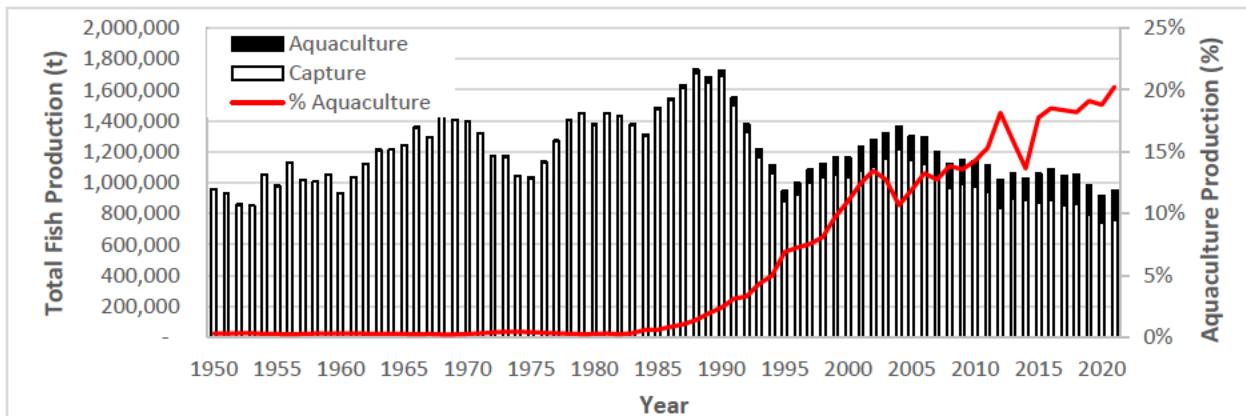


Figure 1. Time series of total Canadian aquatic production from capture fisheries and aquaculture. The line represents the annual percentage of aquaculture contribution to the total production. Data are from FAO. 2023. Fishery and Aquaculture Statistics. Global capture production 1950-2021 (FishStat). In: FAO Fisheries and Aquaculture Division [online]. Rome. Updated 2023. [www.fao.org/fishery/en/statistics/software/fishstati](http://www.fao.org/fishery/en/statistics/software/fishstati)

still dropping<sup>2</sup>. Within Canada, the majority of the seafood production still comes from harvest fisheries (80%). Any significant aquaculture production did not occur until the early 1990s (Fig. 1), although there was some farming for trout and oysters in a few local areas. Since that time, fishery landings have continually dropped by approximately 50% and aquaculture has grown to represent 20% of the roughly 1 million tons of seafood produced in Canada in 2021<sup>3</sup>. The value of the entire Canadian wild fisheries in 2021 was approximately \$4.6 billion<sup>3</sup>, of which the American lobster (*Homarus americanus*) on the east coast represented 44.1% of the total value. Comparatively, the value of the Canadian aquaculture industry in 2021 was approximately \$1.3 billion, of which farmed Atlantic salmon from the east coast represented 22.8% of the total value. Overall, in Canada, salmon represent approximately 75% of the value of the aquaculture industry and 63% of the volume<sup>3</sup>.

4. The lobster fishery on the east coast of North America has experienced some dramatic increases in landings since 1990 with landings increasing over 150%, along with the associated economic benefits to the local economies (Fig. 2). The dominant fishery is in the Gulf of Maine where fishers from both the USA and Canada (primarily Nova Scotia) catch and land lobsters, but landings are also significant for New Brunswick in the Bay of Fundy and Quebec<sup>2,3</sup>. In New England, the recent increase in lobster landings created such a demand for lobster bait (herring primarily) that there were shortages and it was known as the “bait crisis”<sup>4,5</sup>. However, there are now potentially significant changes happening in the overall lobster fishery as the effects of climate change begin to show, changing distributions and challenging management strategies<sup>5-12</sup>.

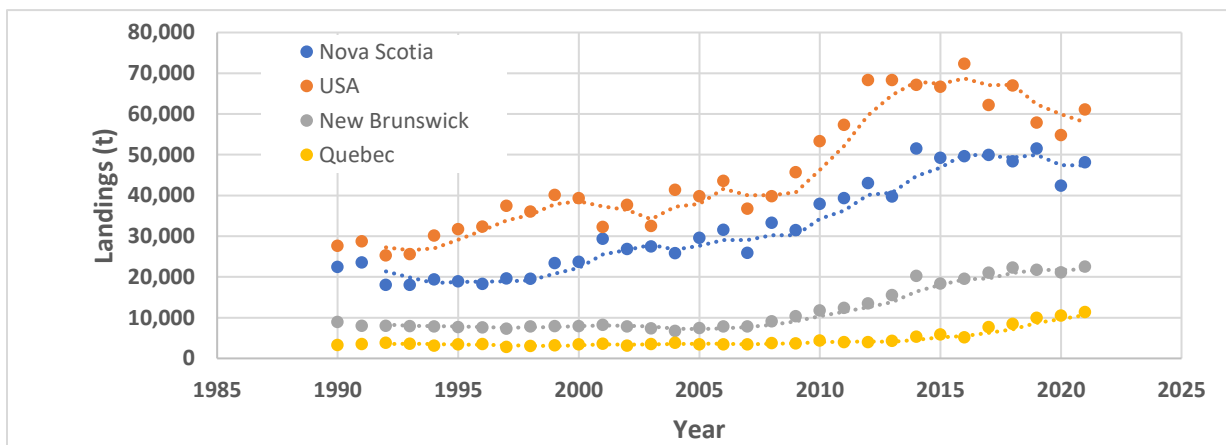


Figure 2. Lobster landings over time from the Gulf of Maine and Gulf of St. Lawrence geographic regions. Fitted lines are 5-point running means. Data are from DFO Statistics Division (<https://www.dfo-mpo.gc.ca/stats/commercial/sea-maritimes-eng.html>.)

5. Lobsters have begun to move north away from the southerly warming waters and as a result, are much less available to the fishers in the southern Gulf of Maine<sup>8</sup>. The peaking of landings and slow decline from the USA landings are readily apparent in Figure 2. Interestingly, there also appears to be a plateauing in the Nova Scotia landings possibly indicating a potential upcoming drop similar to that experienced by fishers in the USA, although unfortunately, the data only go up to 2021 as the most recent landings have not yet been posted publicly. However, word-of-mouth from fishers in the Yarmouth area in December 2023 suggest that opening day lobster landings are down significantly from previous years (Robinson personal observation).
  
6. In comparison to lobsters, the Atlantic salmon aquaculture industry in Canada, and globally, is a relative newcomer to the marine food economy. Starting in the mid-1980's, the salmon aquaculture industry has grown steadily and produces about 3 million tonnes of product a year (2021)<sup>2</sup>, which highlights the growing acceptance and demand for their product (Fig. 3).

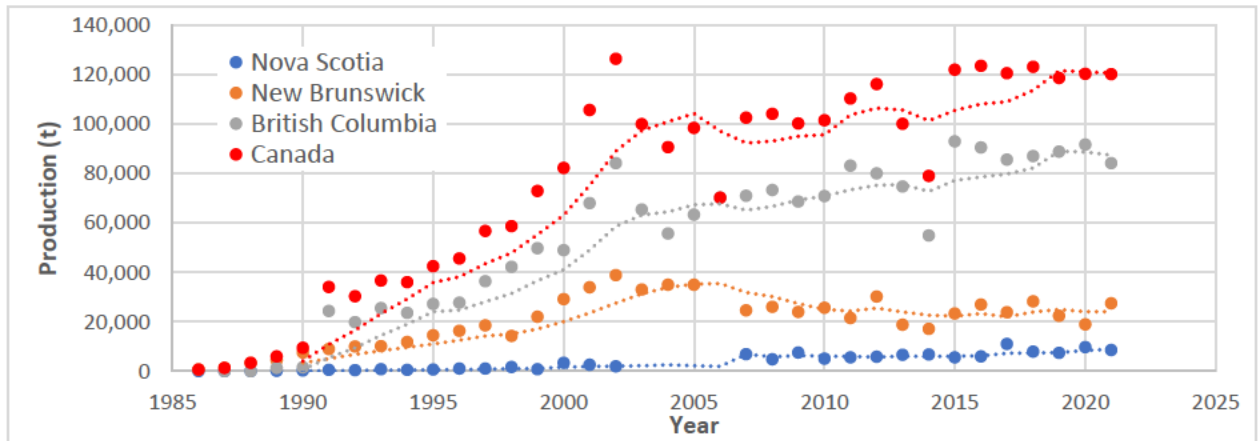


Figure 3. Salmon production over time broken down by province. Fitted lines are 5-point running means. Data are from DFO Statistics Division (<https://www.dfo-mpo.gc.ca/stats/commercial/sea-maritimes-eng.html>.)

7. Canada produces about 120,000 tonnes of salmon annually, as of 2021<sup>3</sup>. British Columbia produces the major share of the fish (70%) followed by New Brunswick (22%), and Nova Scotia (7%). In comparison to other Atlantic salmon producing areas of the world, Canada is the 4<sup>th</sup> largest grower, but only produces 4% of the farmed salmon based on records from 2021; about the same as the Faroe Islands<sup>2</sup>. Norway and Chile dominate the market.

8. With the overall value and employment represented by lobsters and salmon to the coastal economies of the Maritimes, it is imperative to the local communities, who derive the benefits, that both of these industries coexist with each other, ideally with no detrimental effects to either operation. Determining and managing the interactions and the effects between the two industries is complicated as the lobster fishery is more dispersed in its area of operation while the salmon farming industry is more concentrated in space; both using totally different approaches to the production of seafood. How lobsters might be impacted by these seafood-producing industries creates even more uncertainty as lobsters are notoriously difficult to study in the wild since they live in relatively deep water which affects direct observation, are long-lived, are seasonally mobile, act as refuging predators that can spend part of their time hidden under rocks and are most active at night<sup>13</sup>.
9. The goal of my review is to further contribute to the information available on what we know about the interactions between lobsters and Atlantic salmon farming for the purpose of assessing the risk associated with the expansion of the salmon aquaculture industry in Nova Scotia for the use of industry, public managers, the other area users and the salmon farming industry itself. There have been other broader summaries compiled that review the general lobster-salmon farming interactions<sup>14</sup> and also assess the risk with the proposed expansion of the salmon farming infrastructure in Liverpool Bay<sup>15</sup>. I hope that this report will complement some of that information, based on my research experiences in the Bay of Fundy and Atlantic coast over the last 3 decades.
10. There are a number of categories of potential interactions between fish farms and lobsters. They are:
  - (a) Biological;
  - (b) Environmental;
  - (c) Animal health (minimal review in this report); and
  - (d) Genetic (not reviewed in this report).



## Biological Interactions

11. Considering the value of the Maritime lobster fishery in southwest Nova Scotia, it is surprising there are not more biological/ecological studies done in this area. Within the Liverpool Bay area under discussion, there are relatively few direct observational studies done on the lobsters. The most recent studies in the near-region (Port Mouton) have collaborated with the local lobster fishers and used their catch statistics to infer some of the dynamics that may be occurring between the salmon farming and lobster fishing industries<sup>16-18</sup>. Some of the published papers from these studies reported the subjective observations made by lobster fishers during the development of the salmon farming industry<sup>18</sup> while others analyzed the catch-per-unit effort (kg of lobsters per trap haul)<sup>16,17</sup>. The conclusions from these studies were that salmon farms had a negative effect on lobsters and drove larger commercial-sized lobsters away from the farm area as well as reproductively mature females with eggs (berried lobsters). The authors also felt there was a weak interaction with water temperature, but the dataset used was not extensive enough to make any significant conclusions. Up until 2019, these were the best data available in Nova Scotia (11 years of catch observations) to judge the effect of salmon farming on lobster behaviour, even though the conclusions were mostly based on correlation. There was some criticism published in the literature on the conclusions from these studies<sup>19</sup>, but most of the objections revolved around the study design and the extrapolation of trends in the dataset. Needless to say, the original authors disagreed<sup>20</sup>.
12. In New Brunswick, in response to similar concerns about the potential negative interactions that may arise from salmon farms on lobsters, an 8-year study was initiated (2008-2015) on Grand Manan to examine changes in lobster abundance over time. Unlike the Nova Scotia studies which were based on commercial trapping records, the New Brunswick study used diver-based surveys of lobsters at the farm and reference sites during the summer months when the lobster abundance was highest and prior to their seasonal migration into deeper water during the fall and winter months<sup>21</sup>. The conclusions of this study were that there were no detectable interactions between the salmon farm and the local lobster populations or the associated fishery.
13. The above regional studies were both ambitious and commendable for the effort and costs that were involved in carrying them out. Working in the field is almost always exceedingly difficult to generate extensive and representative data sets, particularly when it involves

boats, people and limited financial resources and time periods. Both of these studies were based on catch-per-unit-effort (CPUE) approaches, one with lobster traps and the other visually with divers. Two analytical obstacles that both studies had to overcome were: 1) the uniform catchability (known as  $q$  in the fishery theory) of the lobsters among the different study areas and 2) the ability to detect a subtle signal within a larger external one (rapidly increasing landings). CPUE studies are known to be problematic in determining the state of fish populations as the probability of capturing an animal has to remain the same across all the sampling stations and times<sup>22</sup>. If the catchability is not the same, then the conclusions may not accurately reflect the reality of the changes in population densities. In lobsters in southwestern Nova Scotia, it has been shown that catchability with traps does not remain constant over the season or even with habitat (boulders vs. low relief areas)<sup>23</sup>. In Port Mouton and Grand Manan, the zones in the study area were different in relation to the benthic habitat, so this was an element of variability that was not accounted for. The other issue regarding the conclusions of the studies was that the work was conducted during a time when the lobster population was dramatically increasing and fishery landings continued to increase year after year setting new historical records. If the signal for the interaction between salmon farms and the lobsters was subtle, it may never have been detectable within the larger variability associated with the overall lobster population increases. For example, the lobster catch rates in the Port Mouton study were about 1 Kg/trap haul in the early spring as the lobsters were moving into shallower water<sup>17</sup>. This is about an order of magnitude lower than some of the landings that were achieved in 2019 during our study in Liverpool (see below). This does not mean that the Port Mouton data were flawed, but that the differences in catch rates could be related to either lobster density, catchability or a combination of both, making the subsequent interpretation difficult on top of the very strong signal of a regionally expanding lobster population.

14. Obviously, the interaction between wild species and aquaculture operations is a complex one and one that is of interest to resource managers, both domestically and internationally, as aquaculture expands. There have been several reviews done on the subject that show that there is both an attractive and a repulsive nature to farms<sup>24</sup>. As to be expected, the answers are not simple and depend on the magnitude of the stressor, the time frame, susceptibility of the organisms etc. Short-term predictions are much easier than longer term ones due to the complexity of the interactions and modelling is often used as a tool for assessing potential impacts and scales<sup>25-29</sup> with the large amounts of data to be

interpreted. However, models are only as good as the data used to create them and therefore empirical information is still required to answer management questions.

15. For lobsters, the current pressing management issue is whether salmon farms repel lobsters, based on the CPUE work (above). To answer this question, a new approach was required to generate some information. Technology has provided some interesting options as electronic miniaturization of tracking tags has permitted scientists to use them to unobtrusively follow animal movements over time and space. This is an active and growing field of research for marine animals including crustaceans<sup>30</sup>. Basically, a transmitter is attached to an animal that transmits a signal (e.g. a few minutes to hours) that can be detected by a grid of receivers (essentially microphones). These receivers will then triangulate the signal and determine the position, just like cell phones on a tower network. This provides a long-term record of the position of the animal from which interpretations of the behaviour can then be determined. In 2014, the DFO McKindsey team initiated tracking studies on lobsters and rock crabs in relation mussel aquaculture operations in the Gulf of St. Lawrence on the Îles de-la-Madeleine<sup>31</sup> and Prince Edward Island<sup>32</sup>. The results from these studies unequivocally demonstrated the usefulness of this approach and demonstrated that lobsters indeed used the mussel farms to forage, but remained quite mobile and therefore, were still available to the lobster fishery.
16. In 2016, this same approach was adopted to look at the interactions between lobsters and salmon aquaculture farms in southwest New Brunswick. Study sites were set up with an array of receivers in the Quoddy region at three salmon farms and lobsters were tagged and released to follow their movements through the summer and fall. Like the Gulf of St. Lawrence lobsters, the lobsters readily moved under the salmon farms with the exception of one farm in Back Bay. The time that a lobster spent under the farm seem to be related to shelter habitat since those farms with rocky bottoms had lobsters that remained in place for many weeks making regular foraging trips out and then returning to their burrow. Rock crabs were also very attracted to the areas underneath the farms, but are also prey items for the lobster<sup>33</sup> resulting in interesting trophic food web dynamics.
17. In 2019, the lobster tagging study was extended to Nova Scotia at the invitation the NS Dept. Fisheries and Aquaculture to look at lobster-salmon farming interactions in Liverpool Bay using the techniques developed in New Brunswick and the Gulf of St. Lawrence. A grid array of receivers was established in Liverpool Bay in 2019, 2020 and 2021. Lobsters

were tagged with transmitters and released at the existing farm as well as the two potential farm sites in Fralick Cove (also known as Brooklyn) and Mersey Point. The receivers were deployed in May and were retrieved in November for each year in order not to interfere with the lobster fishing season. The lobster movement results from this study were consistent with the previous studies in New Brunswick which demonstrated that lobsters were very mobile and would move in and around the Coffin Island salmon farming site during their movements around Liverpool Bay with no obvious aversion to the farm. The operating farm near Coffin Island was primarily a rippled sand bottom with no shelter for lobsters, so similar to the New Brunswick observations, the lobsters moved in and out of the farm area during their foraging activities. The proportion of tagged lobsters that frequented the salmon farm decreased from 2019 (fallow year) to 2021 (second year of production), but the lobsters still ventured under the farm. During the course of the 3-year study, 6 egg-bearing female lobsters (berried) were tagged with acoustic transmitters. Two of them were tagged adjacent to the salmon farm in 2020 and either made a few excursions underneath or remained in the vicinity. Crabs, on the other hand, were consistently attracted to the area under the cages during all three study years 2019-2021 and remained there for much longer periods which was similar to the observations in New Brunswick.

18. During the 2020 and 2021 September sampling trips, underwater videos were taken with a remote operated vehicle (ROV - DeepTrekker™) of the sea bottom for the three study sites in Liverpool Bay. At each site, the ROV was lowered over the side of the boat where the operator piloted it to the bottom and attempted to survey along a particular heading, depending on the drift from the wind and the tide which could override the thrust from the small propellers. Video footage was captured on an SD card in the surface controller. Results from the video footage (see attached USB of the video footage at **Tab 1**) showed that the benthic area under the cages of the active Coffin Island salmon farm was a firm rippled sand with many lobsters and crabs moving around and creating shallow depression burrows. The Fralick Cove site (Brooklyn) was mostly rock and ledge covered with seaweed (poor coverage due to entanglement with seaweed) and the Mersey Point site which was a combination of sand and boulder field with a low algal turf. These habitat observations were consistent with a previous study that had surveyed the entire bay with remote sensing technology to classify benthic habitat types for marine spatial planning for lobster management<sup>34</sup>. Interestingly, no lobsters were seen in the Mersey Point and

Fralick Cove (Brooklyn) reference areas from the brief video footage, although these areas are fished regularly during the fishing season and we caught lobsters easily in traps from both areas when we were sampling for the microbiome study (below). This is a good example of the “catchability” issue from an observational point of view when you are looking at complex versus simple habitats as discussed above.

19. My conclusions from the telemetered tagging studies were that lobsters were not actively repelled by salmon farms as suggested by previous studies and that lobsters likely use the areas for foraging, possibly on the crabs that seem to actively inhabit the area, although the use by tagged lobsters appeared to decline as production on the farm grows. The residence time of the lobsters under a salmon farm may be related to habitat availability as sand or mud sites showed that lobsters spent far less time there. Lobsters may not need to range as far inside the farm over the production cycle as the crab population on the lease is quite abundant.
20. The successful foraging of lobsters on food derived from salmon farms was supported by a study in Grand Manan, New Brunswick that looked at the biochemical composition of the crabs and lobsters captured under the farm<sup>35</sup>. That study utilized fats only found in salmon feed as a tracer to show that the lobsters and crabs were obtaining some of the nutrients from the fish food, either from direct feeding or secondarily from other prey species. These results were also confirmed in an unrelated study in Ireland looking at the European lobster on the effects of salmon farms on the local crustaceans<sup>36</sup>.
21. As a result of the new information from the telemetered tagging work and the biochemical analysis, it became apparent that lobsters and crabs were associating with salmon aquaculture sites and also deriving a benefit from them. The next obvious question was: are the lobsters benefiting from the additional food that is becoming available to them or are there some chronic effects that might affect the lobster population in the long term, through physiological changes to growth, reproduction or survival.
22. Determining physiological changes in wild populations in nature is quite problematic as it is impossible to control all the variables affecting the organisms under study. This is similar to the problems faced with using CPUE to determine population effects of a particular stressor. The variation introduced by seasonal, interannual and other outside factors make the evaluation very difficult. For example, if we looked at fecundity of the lobster

population in relation to aquaculture, we would only generate one data point a year since the lobsters only reproduce seasonally. It would take many years to gather enough data to make any reliable assessment of the effects of the culture activities on the egg production of females. The same problem is associated with growth measurements as lobsters grow by moulting their shell and therefore only increase in size in seasonal increments.

23. Since we only had 3 years to conduct the study (2019-2021), we decided to use an integrative measure that has been shown to be indicative of an organism's health. With the development of new technology for the DNA-based study of bacteria, there has been a massive amount of research done in the last decade on the relationship between the population biodiversity of bacteria in the gut of an organism (known as the microbiome) and the organisms' overall health<sup>37-42</sup>. Today, there are over 24,000 papers a year being published on the role of microbiomes<sup>43</sup> in humans, other organisms and the environment. While the majority of publications are in the human medicine category, there is a rapidly expanding database on fish and invertebrates that are showing the same type of relationships between diet and environment on the gut microbiomes of the animal and the overall effect on the health and fitness of the animal<sup>44</sup>. The approach of investigating the gut microbiome in crustaceans as an indicator of overall fitness is also advancing quickly and has been used a number of different lobster species in a wide range of environments<sup>45-53</sup>.
24. Therefore, a study was initiated in 2019 to study the gut microbiome in lobsters from Liverpool Bay at the three different study sites and at Port Mouton (2019 only). Sampling was done for 3 years (2019-2021) in the middle of September for the sake of consistency, although extra samples were also taken in July 2021 to test for any effects of seasonality. Lobsters were captured with commercial lobster traps baited with herring and were set for 24 hours. Sediment samples were also taken at each site where the trapping occurred and were frozen at -80 °C. Traps were retrieved after 24 hours and lobsters were randomly sampled, placed on ice and returned to the lab where they were euthanized and their stomachs removed and frozen immediately at -80 °C. Samples were sent to the Research Productivity Council in Fredericton, New Brunswick where all the samples were extracted for DNA, the rRNA genes amplified, sequenced and then curated to determine bacterial species diversity.

25. This approach of looking at gut microbiomes generated very large datasets and samples would contain hundreds of genera to analyze. The DNA-based taxonomic results clearly showed differences among the lobsters, crabs and the sediment (Fig. 4). Note that each group (crab, lobster and the sediment) clustered together with very little overlap. These clusters were significantly different from each other indicating that the lobsters were maintaining their own unique microbiome compared to the crabs and the environment in which they lived.

26. The data sets also showed interannual variability and indicated that each year had to be considered separately rather than lumping them all together. In the last year (2021), the comparison between samples taken in July and those in September were also different indicating that there is a

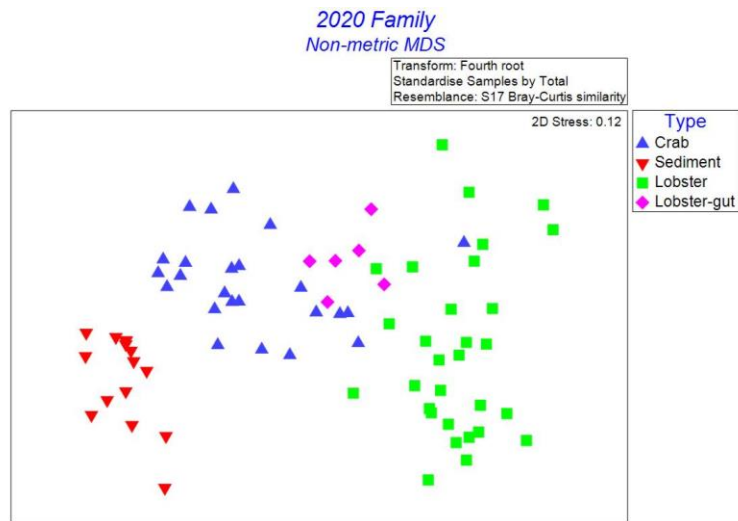


Figure 4. Plot showing the grouping of the microbiomes for crab and lobster stomachs, lobster intestine and the surficial sediment microbiome in the three study locations. Note how each of the groups tend to separate out. The data were produced using a non-metric multidimensional space analysis in the software Primer 7.

seasonal succession of gut and benthic bacterial species. This is not surprising as it has been found in previous planktonic and benthic microbial studies<sup>54-57</sup>.

27. Most importantly, there were no significant differences in the gut microbiomes found in the lobsters from the reference areas and the lobsters from the farm site (Fig. 5) even though the microbiome of the sediment in the Coffin Island salmon farm was different than the ones in Mersey Point of Fralick Cove (Brooklyn) (not shown). These results do not mean that there were no effects on the overall condition of the lobsters from the farm site, but based on the microbiome biodiversity approach with the study of lobster stomachs, we could not detect any effect. The lack of a significant signal within the lobster stomachs would suggest that the effect is too small to detect, if it is there at all. Another approach



may be warranted, but this technique has been shown to produce results in other lobster impact studies<sup>49-51</sup>.

28. Overall, the tagging and the microbiome studies from the Liverpool area show that there are very few detectable negative effects of the farm on the local lobster populations. Lobsters will freely range under and around fish farms and will actively consume some of the nutrients coming from them. This consumption of food items does not seem to be reflected in the microbiome of the lobster stomachs and therefore, there is no observable signal that the gut physiology of the lobsters is being affected. Interestingly, the crabs that actively remained and foraged under the salmon farm did show a change of microbiome between farm and reference sites (Fig.6). This observation would appear to support

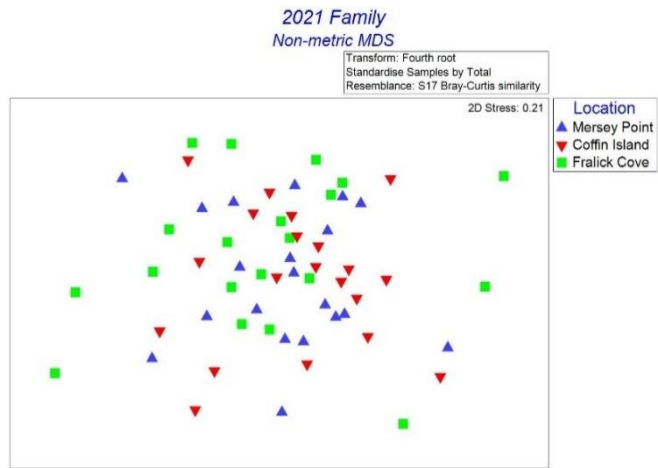


Figure 5. Plot showing the grouping of the microbiomes for lobsters found at the Coffin Island salmon farm and the other two references areas (Fralick Cove (Brooklyn) and Mersey Point). Note the lack of distinct clustering among the three sites.

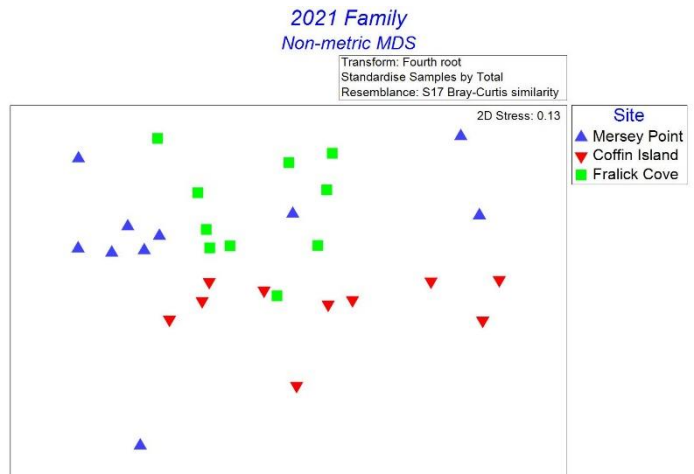


Figure 6. Plot showing the grouping of the microbiomes for rock crabs found at the Coffin Island salmon farm and the other two references areas (Fralick Cove (Brooklyn) and Mersey Point). Note the grouping of the salmon farm site.

the technique to assess the impact on lobster populations.



## Environmental Interactions

29. There have been a number of reviews on the effects of salmon farming on the environment and associated species in the area<sup>14,24,58-62</sup> and it is reasonably well established that the addition of new organic food sources and additional 3-dimensional structures to the environment changes the dynamics of the original ecosystem. This is the same for any anthropogenic impact in the marine ecosystem, including mining, oil and gas, farming, fishing, wastewater treatment and coastal zone development. Since it is virtually impossible to reduce the impacts of industrial use to zero, management objectives have centered on monitoring the effects and creating policies that will minimize the impact to the local environment based on a risk assessment to existing resources. For aquaculture, this has led to the creation of the federal Aquaculture Activity Regulations<sup>63</sup> as well as policies and regulations that have been created at the provincial level, where the administration of the industry occurs, to manage aquaculture development and operations. Science at the federal, provincial and academic level contributes to the policies through direct research on issues as well as review of the literature to create summaries of interactions, such as pathways of effects<sup>64</sup> and better ways to incorporate local knowledge from the communities<sup>65</sup>.
30. Monitoring of the benthic organic impacts of salmon farming in Canada are primarily done through measuring the free sulfide levels in the sediment<sup>66</sup> (caused by benthic loading of organic material) and the infaunal biodiversity. Sampling is done with small vessels around the farms, often by monitoring companies on the east coast and DFO staff on the west coast. Sample processing for biodiversity in the sediments is slow and can often take trained taxonomists considerable lengths of time to accomplish. Monitoring methods for sulfide levels is changing and new approaches are being adopted to increase the accuracy and speed of sampling<sup>67,68</sup>.
31. Internationally, there are new methods for monitoring environmental changes that are increasing the resolution of management. The conditions of any habitat will determine the types of species that can exist there and this relationship exists at the macrolevel with larger organisms to the micro-level of bacteria, viruses etc. Bacterial microbiomes have been extensively studied and shown that their biodiversity and changes over time can reflect impacts to the ecosystem<sup>69-72</sup>. Since the reproductive time of bacteria are measured in hours or a few days, changes to the microbiome can happen reasonably

quickly and give a more sensitive estimate of environmental changes. This technique is currently being considered or being applied in New Zealand, Scotland and Norway for salmon aquaculture monitoring<sup>73-77</sup>.

## **SUMMARY STATEMENT**

32. My overall conclusion from a review of the literature on field-based studies on lobsters and the recently collected data from the lobster acoustic tracking and microbiome projects is that salmon farms in Liverpool Bay will have little negative effect on the behaviour and distribution of lobsters that could affect the local fishery.
33. Previous studies using the best available data at the time were hampered in their conclusions with the assumptions they had to make on the consistency of catch rates within their studies (CPUE). Our direct observational field study using acoustic tracking technology demonstrates that local rock crab populations do capitalize on the increase in organic material coming from the farms as their distribution and abundance increase in the vicinity of the salmon farms whereas lobsters are attracted, but not as intensively as crabs. Tracers such as fatty acids, and now bacterial microbiomes from the stomach, confirm that there is a pathway of effect to the crabs. Crabs are a known prey item for lobsters and the tagging and ROV video clearly show that lobsters do forage under the salmon farms, which refutes the conclusions of the previous studies. While the lobsters also show a biochemical signal (fatty acids) from the farm in our previous study, there is no sign that the organic output from the farm is changing the microbiome biodiversity in the stomachs of the lobster.

## **Future suggestions with respect to monitoring Liverpool Bay**

### **Rationale**

34. It is highly likely that near-future physical and biological changes will occur in the marine environment of Liverpool Bay due to climate change and these will very likely affect both commercial populations of lobsters and the salmon farming industry. The salmon farming industry is already planning on doing enhanced environmental monitoring. Warming waters, changes in hydrographic conditions and changing distributions of animals, seaweeds and microorganisms will all combine to create more variability, uncertainty and therefore more difficulty in managing the bay. As such, future studies, led by provincial

and university scientists, and supported by the industry sector, could perhaps use the Liverpool Bay farm expansion as a long-term case study area, addressing concerns. A monitoring program would have to be well-designed and implemented by the NS Dept. Fisheries and Aquaculture to detect changes within the larger upcoming climate-change signal. Long term field-based research on impacts of climate change and aquaculture activity are particularly scant in the literature.

### **Who should be involved?**

35. The Province of Nova Scotia should be involved as they regulate the industry provincially as well as running the current environmental monitoring program. They also have significant logistic resources that can be mobilized in both people and equipment.
36. DFO and Environment Canada and Climate Change (ECCC) could be involved to provide answers to regulatory science and climate change issues since they have the national mandate for these files. They also have logistic resources that can be mobilized.
37. Academia should be involved to study the biological and physical processes that will be occurring in Liverpool Bay and around the salmon farms. For example, this could involve changes to population biology parameters such as growth, reproduction and recruitment or it may involve changes to pathogen pressures on farmed and wild fish.
38. Industry should be involved through providing logistic support for monitoring where appropriate. This could involve the use of vessels, staff etc. as appropriate.

### **What should be monitored?**

39. I would suggest that the traditional monitoring protocols be expanded as they will likely not be enough to monitor some of the more subtle potential impacts of climate change and the more minor signal from new salmon farms on local lobster populations. What to monitor should be determined jointly by all the users of Liverpool Bay. Carefully crafted questions need to be developed that can be scientifically tested over a specific length of time, depending on the question. That could be either a few years or perhaps over a decade or more.

### **How can this be achieved?**

40. Coordination of this monitoring program should be handled by the Province of Nova Scotia as they currently run the environmental monitoring program. Federal agencies (DFO, ECCC) have access to larger national programs for funding and have the mandate for managing commercial fishing populations and initiatives related to climate change.
41. The current suite of monitoring protocols should be continued, but newer approaches should also be employed that can be evaluated for the rest of the province and possibly the country. Examples of this would be some of the genetic technologies such as metagenomics to monitor environmental impacts and the use of environmental DNA (eDNA) to count organisms. This type of technology is already being developed in other countries so a technology transfer would be very valuable and save development time.

## References

- 1 Lear, W. H. History of fisheries in the Northwest Atlantic: the 500-year perspective. *Journal of Northwest Atlantic Fishery Science* **23** (1998).
- 2 FAO. *Fishery and Aquaculture Statistics. Global capture production 1950-2021 (FishStatJ)* [www.fao.org/fishery/en/statistics/software/fishstatj](http://www.fao.org/fishery/en/statistics/software/fishstatj), 2023).
- 3 DFO. *Statistics* (<https://www.dfo-mpo.gc.ca/stats/stats-eng.htm>), 2023).
- 4 Grabowski, J. H. *et al.* Use of herring bait to farm lobsters in the Gulf of Maine. *PLoS One* **5**, e10188, doi:10.1371/journal.pone.0010188 (2010).
- 5 Stoll, J. S. *et al.* Rapid adaptation to crisis events: Insights from the bait crisis in the Maine lobster fishery. *Ambio* **51**, 926-942, doi:10.1007/s13280-021-01617-8 (2022).
- 6 Steneck, R. S. *et al.* Creation of a gilded trap by the high economic value of the Maine lobster fishery. *Conserv Biol* **25**, 904-912, doi:10.1111/j.1523-1739.2011.01717.x (2011).
- 7 Schultz, L., Folke, C., Österblom, H. & Olsson, P. Adaptive governance, ecosystem management, and natural capital. *Proc Natl Acad Sci U S A* **112**, 7369-7374, doi:10.1073/pnas.1406493112 (2015).
- 8 Le Bris, A. *et al.* Climate vulnerability and resilience in the most valuable North American fishery. *Proc Natl Acad Sci U S A* **115**, 1831-1836, doi:10.1073/pnas.1711122115 (2018).
- 9 Waring, T. & Acheson, J. Evidence of cultural group selection in territorial lobstering in Maine. *Sustain Sci* **13**, 21-34, doi:10.1007/s11625-017-0501-x (2018).
- 10 Oppenheim, N. G., Wahle, R. A., Brady, D. C., Goode, A. G. & Pershing, A. J. The cresting wave: larval settlement and ocean temperatures predict change in the American lobster harvest. *Ecol Appl* **29**, e02006, doi:10.1002/eap.2006 (2019).
- 11 Staudinger, M. D. *et al.* It's about time: A synthesis of changing phenology in the Gulf of Maine ecosystem. *Fish Oceanogr* **28**, 532-566, doi:10.1111/fog.12429 (2019).
- 12 McClenachan, L., Scyphers, S. & Grabowski, J. H. Views from the dock: Warming waters, adaptation, and the future of Maine's lobster fishery. *Ambio* **49**, 144-155, doi:10.1007/s13280-019-01156-3 (2020).
- 13 Karnofsky, E. B., Atema, J. & Elgin, R. H. Natural dynamics of population structure and habitat use of the lobster, *Homarus americanus*, in a shallow cove. *Biol Bull* **176**, 247-256, doi:10.2307/1541983 (1989).
- 14 Horricks, R. A., Lewis-McCrea, L. M. & Reid, G. K. Interactions between American lobster (*Homarus americanus*) and salmonid aquaculture in the Canadian Maritimes. *Canadian Journal of Fisheries and Aquatic Sciences* **79**, 1561-1571, doi:10.1139/cjfas-2021-0252 (2022).
- 15 DFO. DFO Maritimes Region Science Review of the Proposed Marine Finfish Aquaculture Boundary Amendment and New Sites, Liverpool Bay, Queens County, Nova Scotia. . *DFO Can. Sci. Advis. Sec. Sci. Resp.* **2022/039.**, 1-56 (2022).
- 16 Loucks, R. H., Smith, R. E. & Fisher, E. B. Interactions between finfish aquaculture and lobster catches in a sheltered bay. *Marine Pollution Bulletin* **88**, 255-259, doi:10.1016/j.marpolbul.2014.08.035 (2014).
- 17 Milewski, I. *et al.* Sea-cage aquaculture impacts market and berried lobster (*Homarus americanus*) catches. *Marine Ecology Progress Series* **598**, 85-97 (2018).
- 18 Wiber, M. G., Young, S. & Wilson, L. Impact of Aquaculture on Commercial Fisheries: Fishermen's Local Ecological Knowledge. *Human Ecology* **40**, 29-40, doi:10.1007/s10745-011-9450-7 (2012).
- 19 Grant, J., Filgueira, R. & Barrell, J. Lack of interaction between finfish aquaculture and lobster catch in coastal Nova Scotia. *Marine Pollution Bulletin* **110**, 613-615, doi:10.1016/j.marpolbul.2016.06.043 (2016).

- 20 Loucks, R. H., Smith, R. E. & Fisher, E. B. A Response to the letter to the editor 'Lack of interaction between finfish aquaculture and lobster catches in coastal Nova Scotia'. *Marine Pollution Bulletin* **110**, 616-618, doi:10.1016/j.marpolbul.2016.06.111 (2016).
- 21 Grant, J., Simone, M. & Daggett, T. Long-term studies of lobster abundance at a salmon aquaculture site, eastern Canada. *Canadian Journal of Fisheries and Aquatic Sciences* **76**, 1096-1102, doi:10.1139/cjfas-2017-0547 (2019).
- 22 Maunder, M. N. *et al.* Interpreting catch per unit effort data to assess the status of individual stocks and communities. *ICES Journal of Marine Science* **63**, 1373-1385, doi:10.1016/j.icesjms.2006.05.008 (2006).
- 23 Tremblay, M. J. & Smith, S. J. Lobster (*Homarus americanus*) catchability in different habitats in late spring and early fall. *Marine and Freshwater Research* **52**, 1321-1331, doi:<https://doi.org/10.1071/MF01171> (2002).
- 24 Callier, M. D. *et al.* Attraction and repulsion of mobile wild organisms to finfish and shellfish aquaculture: a review. *Reviews in Aquaculture* **10**, 924-949, doi:<https://doi.org/10.1111/raq.12208> (2018).
- 25 Ferreira, J. G. ECOWIN — an object-oriented ecological model for aquatic ecosystems. *Ecological Modelling* **79**, 21-34, doi:[https://doi.org/10.1016/0304-3800\(94\)00033-E](https://doi.org/10.1016/0304-3800(94)00033-E) (1995).
- 26 Nunes, J. P. *et al.* A model for sustainable management of shellfish polyculture in coastal bays. *Aquaculture* **219**, 257-277, doi:[https://doi.org/10.1016/S0044-8486\(02\)00398-8](https://doi.org/10.1016/S0044-8486(02)00398-8) (2003).
- 27 Ferreira, J. G., Hawkins, A. J. S. & Bricker, S. B. Management of productivity, environmental effects and profitability of shellfish aquaculture — the Farm Aquaculture Resource Management (FARM) model. *Aquaculture* **264**, 160-174, doi:<https://doi.org/10.1016/j.aquaculture.2006.12.017> (2007).
- 28 Ferreira, J. G., Saurel, C., Lencart e Silva, J. D., Nunes, J. P. & Vazquez, F. Modelling of interactions between inshore and offshore aquaculture. *Aquaculture* **426-427**, 154-164, doi:<https://doi.org/10.1016/j.aquaculture.2014.01.030> (2014).
- 29 Ferreira, J. G. *et al.* An integrated model for aquaculture production, pathogen interaction, and environmental effects. *Aquaculture* **536**, 736438, doi:<https://doi.org/10.1016/j.aquaculture.2021.736438> (2021).
- 30 Florko, K. R. N. *et al.* Tracking movements of decapod crustaceans: a review of a half-century of telemetry-based studies. *Marine Ecology Progress Series* **679**, 219-239 (2021).
- 31 Lavoie, M. F. *et al.* Movement of American lobster *Homarus americanus* associated with offshore mussel *Mytilus edulis* aquaculture. *Aquaculture Environment Interactions* **14**, 189-204 (2022).
- 32 Lees, K. J. *et al.* Movement of American lobsters *Homarus americanus* and rock crabs *Cancer irroratus* around mussel farms in Malpeque Bay, Prince Edward Island, Canada. *Aquaculture Environment Interactions* **15**, 179-193 (2023).
- 33 Sardenne, F., Forget, N. & McKindsey, C. W. Contribution of mussel fall-off from aquaculture to wild lobster *Homarus americanus* diets. *Mar Environ Res* **149**, 126-136, doi:10.1016/j.marenvres.2019.06.003 (2019).
- 34 McKee, A., Grant, J. & Barrell, J. Mapping American lobster (*Homarus americanus*) habitat for use in marine spatial planning. *Canadian Journal of Fisheries and Aquatic Sciences* **78**, 704-720, doi:10.1139/cjfas-2020-0051 (2021).
- 35 Sardenne, F., Simard, M., Robinson, S. M. C. & McKindsey, C. W. Consumption of organic wastes from coastal salmon aquaculture by wild decapods. *Sci Total Environ* **711**, 134863, doi:10.1016/j.scitotenv.2019.134863 (2020).



- 36 Baltadakis, A., Casserly, J., Falconer, L., Sprague, M. & Telfer, T. C. European lobsters utilise Atlantic salmon wastes in coastal integrated multi-trophic aquaculture systems. *Aquaculture Environment Interactions* **12**, 485-494, doi:10.3354/aei00378 (2020).
- 37 Siddiqui, R. & Khan, N. A. Microbiome and One Health: Potential of Novel Metabolites from the Gut Microbiome of Unique Species for Human Health. *Microorganisms* **11**, doi:10.3390/microorganisms11020481 (2023).
- 38 Wang, C. *et al.* Gut microbiome-based strategies for host health and disease. *Crit Rev Food Sci Nutr*, 1-16, doi:10.1080/10408398.2023.2176464 (2023).
- 39 Williams, G. M., Tapsell, L. C. & Beck, E. J. Gut health, the microbiome and dietary choices: An exploration of consumer perspectives. *Nutr Diet* **80**, 85-94, doi:10.1111/1747-0080.12769 (2023).
- 40 Wilson, D. R., Binford, L. & Hickson, S. The Gut Microbiome and Mental Health. *J Holist Nurs*, 8980101231170487, doi:10.1177/08980101231170487 (2023).
- 41 Ren, Y. *et al.* Lifestyle patterns influence the composition of the gut microbiome in a healthy Chinese population. *Sci Rep* **13**, 14425, doi:10.1038/s41598-023-41532-4 (2023).
- 42 Armet, A. M. *et al.* Rethinking healthy eating in light of the gut microbiome. *Cell Host Microbe* **30**, 764-785, doi:10.1016/j.chom.2022.04.016 (2022).
- 43 PubMed. *National Center for Biotechnology Information*, <<https://pubmed.ncbi.nlm.nih.gov/?term=microbiomes>> (2023).
- 44 Harris, J. M. The presence, nature, and role of gut microflora in aquatic invertebrates: A synthesis. *Microbial Ecology* **25**, 195-231, doi:10.1007/BF00171889 (1993).
- 45 Meziti, A., Mente, E. & Kormas, K. A. Gut bacteria associated with different diets in reared *Nephrops norvegicus*. *Syst Appl Microbiol* **35**, 473-482, doi:10.1016/j.syapm.2012.07.004 (2012).
- 46 Meziti, A., Ramette, A., Mente, E. & Kormas, K. A. Temporal shifts of the Norway lobster (*Nephrops norvegicus*) gut bacterial communities. *FEMS Microbiol Ecol* **74**, 472-484, doi:10.1111/j.1574-6941.2010.00964.x (2010).
- 47 Ooi, M. C., Goulden, E. F., Smith, G. G. & Bridle, A. R. Haemolymph microbiome of the cultured spiny lobster *Panulirus ornatus* at different temperatures. *Scientific Reports* **9**, doi:10.1038/s41598-019-39149-7 (2019).
- 48 Ooi, M. C., Goulden, E. F., Smith, G. G., Nowak, B. F. & Bridle, A. R. Developmental and gut-related changes to microbiomes of the cultured juvenile spiny lobster *Panulirus ornatus*. *FEMS microbiology ecology* **93**, doi:10.1093/femsec/fix159 (2017).
- 49 Holt, C. C., Bass, D., Stentiford, G. D. & van der Giezen, M. Understanding the role of the shrimp gut microbiome in health and disease. *Journal of Invertebrate Pathology*, 107387, doi:<https://doi.org/10.1016/j.jip.2020.107387> (2020).
- 50 Holt, C. C., van der Giezen, M., Daniels, C. L., Stentiford, G. D. & Bass, D. Spatial and temporal axes impact ecology of the gut microbiome in juvenile European lobster (*Homarus gammarus*). *The ISME Journal* **14**, 531-543, doi:10.1038/s41396-019-0546-1 (2020).
- 51 Battison, A. L., Després, B. M. & Greenwood, S. J. Ulcerative enteritis in *Homarus americanus*: Case report and molecular characterization of intestinal aerobic bacteria of apparently healthy lobsters in live storage. *Journal of Invertebrate Pathology* **99**, 129-135, doi:10.1016/j.jip.2008.06.013 (2008).
- 52 Tang, L. *et al.* Temperature potentially induced distinctive flavor of mud crab *Scylla paramamosain* mediated by gut microbiota. *Scientific Reports* **10**, doi:10.1038/s41598-020-60685-0 (2020).
- 53 Feinman, S. G., Martínez, A. U., Bowen, J. L. & Tlusty, M. F. Fine-scale transition to lower bacterial diversity and altered community composition precedes shell disease in

- laboratory-reared juvenile American lobster. *Diseases of Aquatic Organisms* **124**, 41-54, doi:10.3354/dao03111 (2017).
- 54 Bunse, C. & Pinhassi, J. Marine Bacterioplankton Seasonal Succession Dynamics. *Trends in Microbiology* **25**, 494-505, doi:10.1016/j.tim.2016.12.013 (2017).
- 55 Mackey, K. R. M. *et al.* Seasonal Succession and Spatial Patterns of Synechococcus Microdiversity in a Salt Marsh Estuary Revealed through 16S rRNA Gene Oligotyping. *Frontiers in Microbiology* **8**, doi:10.3389/fmicb.2017.01496 (2017).
- 56 Liu, J., Qiao, Y., Xin, Y., Li, Y. & Zhang, X. H. Seasonal Succession and Temperature Response Pattern of a Microbial Community in the Yellow Sea Cold Water Mass. *Appl Environ Microbiol* **88**, e0116922, doi:10.1128/aem.01169-22 (2022).
- 57 von Jackowski, A. *et al.* Variations of microbial communities and substrate regimes in the eastern Fram Strait between summer and fall. *Environ Microbiol* **24**, 4124-4136, doi:10.1111/1462-2920.16036 (2022).
- 58 Holmer, M. Environmental issues of fish farming in offshore waters: perspectives, concerns and research needs. *Aquaculture Environment Interactions* **1**, 57-70 (2010).
- 59 Holmer, M. & Kristensen, E. Impact of marine fish cage farming on metabolism and sulfate reduction of underlying sediments. *Marine Ecology Progress Series* **80**, 191-201 (1992).
- 60 Holmer, M. & Kristensen, E. Seasonality of sulfate reduction and pore water solutes in a marine fish farm sediment: The importance of temperature and sedimentary organic matter. *Biogeochemistry* **32**, 15-39 (1996).
- 61 Buschmann, A. H. *et al.* A review of the impacts of salmonid farming on marine coastal ecosystems in the southeast Pacific. *Ices J Mar Sci* **63**, 1338-1345, doi:10.1016/j.icesjms.2006.04.021 (2006).
- 62 Milewski, I. in *Marine Aquaculture and the Environment: A Meeting for Stakeholders in the Northeast*. (ed M.F. Tlusty, Bengston, D.A., Halvorson, H.O., Oktay, S.D., Pearce, J.B., Rheault, R.B., Jr.) 166–197 (Cape Cod Press, 2001).
- 63 Canada. *Aquaculture Activities Regulations (SOR/2015-177)*, <<https://laws-lois.justice.gc.ca/eng/regulations/SOR-2015-177/index.html>> (2023).
- 64 DFO. Science advice on revisiting Pathways of Effects (PoE) diagrams in support of FFHPP risk assessment. *DFO Can. Sci. Advis. Sec. Sci. Advis. Rep.* **2021/053**. (2021).
- 65 Milewski, I., Smith, R. E. & Lotze, H. K. Interactions between finfish aquaculture and American lobster in Atlantic Canada. *Ocean & Coastal Management* **210**, 105664, doi:<https://doi.org/10.1016/j.ocecoaman.2021.105664> (2021).
- 66 Hargrave, B. T., Holmer, M. & Newcombe, C. P. Towards a classification of organic enrichment in marine sediments based on biogeochemical indicators. *Marine Pollution Bulletin* **56**, 810-824, doi:<https://doi.org/10.1016/j.marpolbul.2008.02.006> (2008).
- 67 Cranford, P., Brager, L., Elvines, D., Wong, D. & Law, B. A revised classification system describing the ecological quality status of organically enriched marine sediments based on total dissolved sulfides. *Marine Pollution Bulletin* **154**, doi:10.1016/j.marpolbul.2020.111088 (2020).
- 68 Cranford, P. J., Brager, L. & Wong, D. A dual indicator approach for monitoring benthic impacts from organic enrichment with test application near Atlantic salmon farms. *Marine Pollution Bulletin* **124**, 258-265, doi:10.1016/j.marpolbul.2017.07.049 (2017).
- 69 Cordier, T. *et al.* Ecosystems monitoring powered by environmental genomics: A review of current strategies with an implementation roadmap. *Molecular Ecology*, doi:10.1111/mec.15472 (2020).
- 70 Cordier, T. *et al.* Supervised machine learning outperforms taxonomy-based environmental DNA metabarcoding applied to biomonitoring. *Molecular Ecology Resources* **18**, 1381-1391, doi:10.1111/1755-0998.12926 (2018).



- 71 Dully, V. *et al.* Comparing sediment preservation methods for genomic biomonitoring of coastal marine ecosystems. *Marine Pollution Bulletin* **173**, 113129, doi:<https://doi.org/10.1016/j.marpolbul.2021.113129> (2021).
- 72 Pawlowski, J. *et al.* Environmental DNA metabarcoding for benthic monitoring: A review of sediment sampling and DNA extraction methods. *Science of The Total Environment*, 151783, doi:<https://doi.org/10.1016/j.scitotenv.2021.151783> (2021).
- 73 Frühe, L. *et al.* Global Trends of Benthic Bacterial Diversity and Community Composition Along Organic Enrichment Gradients of Salmon Farms. *Frontiers in Microbiology* **12**, doi:10.3389/fmicb.2021.637811 (2021).
- 74 Stoeck, T. *et al.* Environmental DNA metabarcoding of benthic bacterial communities indicates the benthic footprint of salmon aquaculture. *Marine Pollution Bulletin* **127**, 139-149, doi:10.1016/j.marpolbul.2017.11.065 (2018).
- 75 Keeley, N. *et al.* Mixed-habitat assimilation of organic waste in coastal environments - It's all about synergy! *Science of the Total Environment* **699**, 15, doi:10.1016/j.scitotenv.2019.134281 (2020).
- 76 Keeley, N., Wood, S. A. & Pochon, X. Development and preliminary validation of a multi-trophic metabarcoding biotic index for monitoring benthic organic enrichment. *Ecological Indicators* **85**, 1044-1057, doi:10.1016/j.ecolind.2017.11.014 (2018).
- 77 Pochon, X., Zaiko, A., Fletcher, L. M., Laroche, O. & Wood, S. A. Wanted dead or alive? Using metabarcoding of environmental DNA and RNA to distinguish living assemblages for biosecurity applications. *PLoS ONE* **12**, doi:10.1371/journal.pone.0187636 (2017).

## **Tab 1**

Video footage of the sea bottom for the three study sites in Liverpool Bay taken in September 2020 and September 2021.

Secure link provided to ARB and parties under separate cover.

**TAB B**

**KCS' Application re AQ#1205X, AQ#1432,  
AQ#1433 in Liverpool Bay, Queens County**

This is Exhibit A referred to in the Affidavit  
of Shawn Robinson, PhD, affirmed virtually  
before me on January 19, 2024.



---

Barrister of the Supreme Court of Nova Scotia

## Shawn M.C. Robinson, Senior Scientist

### *Longline Environment*

London, [REDACTED] United Kingdom

[REDACTED] (Telephone) [REDACTED] (email)

### **Education**

Ph.D. (1984-1988) University of British Columbia, Vancouver, B.C.

MSc. (1979-1983) Simon Fraser University, Burnaby, B.C.

BSc. (Honors) (1976-1979) Acadia University, Wolfville, N.S.

### **Appointments**

Research Scientist, Department of Fisheries and Oceans, St. Andrews, NB, 1988-2022 (retired)

Adjunct Professor, University of New Brunswick, Saint John, NB, 2002-2022

Adjunct Professor, Nova Scotia Agricultural College, Truro, NS, 2000-2004

**Short Resume:** Dr. Robinson worked for 34 years as a research scientist with the Dept. Fisheries and Oceans at the Biological Station in St. Andrews, New Brunswick. He was actively engaged in applied ecological research on marine shellfish species such as blue mussels, sea scallops, sea urchins, soft-shell clams, sea lice and marine bacteria. His research team studied the natural processes by which these animals interact and utilize their environment so that better and more sustainable culture techniques could be developed. One example of this research was the study of an integrated multi-trophic aquaculture (IMTA) program (sometimes known as polyculture) where shellfish and seaweeds were grown in conjunction with other fed species to recycle organic matter and produce a more sustainable and productive system. He also investigated marine microbiomes and how they related to the conversion of organic matter in the environment as a result of human activities and contributed to the fitness of marine organisms. Much of this work involved collaborative projects with industry and academic partners (nationally and internationally) and took a more holistic view of the aquaculture system combining biology, physics, economics, sociology, and government policy. He was a member of the Aquaculture Association of Canada (past president). Publications (185) are in the fields of: stock assessment, subtidal marine ecology, fishery management and aquaculture ecology; mostly associated with invertebrates.

### **Selected Publications and Reports:**

Kennedy, EJ; Robinson, SMC; Parsons, GJ; Castell, JD 1999. Somatic growth trials for juvenile green sea urchins fed prepared and natural diets. Bull. Aquacult. Assoc. Can. 99(4):52-54.

Martin, J.L., F.H. Page, S. Robinson, M.M. LeGresley and M. Ringuette. 1999. Salmonid aquaculture mortalities and a bloom of *Mesodinium rubrum* in Passamaquoddy Bay in 1998. p. 40. In: Ollerhead, J., P.W. Hicklin, P.G. Wells and K. Ramsey (eds). 1999. *Understanding Change in the Bay of Fundy Ecosystem*. Proceedings of the 3<sup>rd</sup> Bay of Fundy Science Workshop, Mount Allison University, Sackville, New Brunswick, April 22-24, 1999. Environment Canada, Atlantic Region Occasional Report No. 12, Environment Canada, Sackville, New Brunswick, 143pp.

Robinson, S.M.C. 2000. Southwestern New Brunswick (LFA 36-38) green sea urchins. DFO Science Stock Status Report C3-49 (2000) 8 p.

- Robinson, S.M.C. 2000. The future of the sea urchin industry: A case study from New Brunswick. *Aquaculture Ireland* Aug/Sept 2000: 13-15
- Robinson, S.M.C., J. Castell, E. Kennedy and L. Peters. 2000. A summary of sea urchin culture at the St. Andrews Biological Station. Available at: <http://crdpm.cus.ca/oursin/> 6p.
- Robinson, S.M.C., S. Bernier and A. MacIntyre 2001. The impact of scallop drags on sea urchin populations and benthos in the Bay of Fundy, Canada. *Hydrobiologia* 465:103-114.
- Kennedy, E., S.M.C. Robinson, G.J. Parsons and J.D. Castell. 2001. Studies on feed formulations to maximise somatic growth rates of juvenile green sea urchins (*Strongylocentrotus droebachiensis*). *Aquacul. Assoc. Canada Spec. Publ* 4:68-71.
- Pearce, C.M., T.L. Daggett and S.M.C. Robinson. 2001. Effect of binder type and concentration on prepared feed stability and gonad yield and quality of the green sea urchin, *Strongylocentrotus droebachiensis*. *Aquaculture* 205:301-323.
- Robinson, S.M.C., J.D. Castell and E.J. Kennedy. 2002. Developing suitable colour in the gonads of cultured green sea urchins (*Strongylocentrotus droebachiensis*). *Aquaculture* 206: 289-303.
- Auffrey, L. and S. Robinson. 2001. Effects of green macroalgal mats on the population dynamics of soft-shell clams (*Mya arenaria*) in southwestern New Brunswick. pp.35-37. In: Hargrave, B.T. and G.A. Phillips (Eds.) *Environmental Studies for Sustainable Aquaculture (ESSA):2001 Workshop Report*. *Can. Tech. Rep. Fish. Aquat. Sci.* 2352:vii + 73 pp.
- Andrew, N., Y. Agatsuma; A. Bazhin, E. Creaser, D. Barnes, L. Botsford, A. Bradbury, A. Campbell, S. Einnarsson, P. Gerring, K. Hebert, M. Hunter, S.B. Hur, C. Johnson, M.A. Juinio-Meñez, P. Kalvass, R. Miller, C. Moreno, J. Palleiro, D. Rivas, S. Robinson, R. Steneck, R. Vadas, D. Woodby and Z. Xiaoqi. 2001. Status and Management of World Sea Urchin Fisheries. *Oceanogr. Mar. Biol. Annu. Rev.* 40: 343-425.
- Pearce, C.M., T.L. Daggett and S.M.C. Robinson. 2002. Optimizing prepared feed ration for gonad production of the green sea urchin, *Strongylocentrotus droebachiensis*. *J. World Aqua. Assoc.* 33: 268-277.
- Chandler, R.A., S.M.C. Robinson and J.D. Martin. 2001. Collection of soft-shell clam (*Mya arenaria* L.) spat with artificial substrates. *Can. Tech. Rep. Fish. Aquat. Sci.* 2390:vii + 11pp.
- Kennedy, E., S.M.C. Robinson, G.J. Parsons and J.D. Castell. 2001. Importance of dietary minerals and pigments in enhancing somatic growth of juvenile green sea urchins (*Strongylocentrotus droebachiensis*). *Aquacul. Assoc. Canada Spec. Publ* 5:31-34.
- Auffrey, L., S.M.C. Robinson and M.A. Barbeau. 2002. Green algae and clams: What's the story? pp.84-91. In: Hargrave, B.T. (Ed.) *Environmental Studies for Sustainable Aquaculture (ESSA):2002 Workshop Report*. *Can. Tech. Rep. Fish. Aquat. Sci.* 2411:v + 117 pp.
- Pearce, C.M., T.L. Daggett and S.M.C. Robinson. 2002. Effect of protein source ratio and protein concentration in prepared feeds on gonad yield and quality of the green sea urchin, *Strongylocentrotus droebachiensis*. *Aquaculture* 214:307-332
- Barber, B., G.S. MacCallum, S.M.C. Robinson, S. McGladdery. 2002. Occurrence and lack of transmissibility of gonadal neoplasia in softshell clams, *Mya arenaria*, in Maine (USA) and Atlantic Canada. *Aquat. Living Resour.* 15(5):319-326.
- Robinson, S.M.C. 2003. Book Review: *Edible Sea Urchins: Biology and Ecology*. *Aquaculture* 217:684-685.
- Haya, K., J.L. Martin, S.M.C. Robinson, J.D. Martin and A. Khots 2003. Does uptake of *Alexandrium fundyense* cysts contribute to the levels of PSP toxin found in the giant scallop, *Placopecten magellanicus*? *Harmful Algae* 2(1): 75 – 81.

- Pearce, C.M., T.L. Daggett and S.M.C. Robinson. 2003. Effects of starch type, macroalgal meal source and  $\beta$ -carotene on gonad yield and quality of the green sea urchin *Strongylocentrotus droebachiensis* (Muller) fed prepared diets. *J. Shellfish Res.* 22(2): 505-520.
- Milligan, T., S.Robinson and P. Yeats. 2003. Particle aggregation and tracers for particulate dispersion from finfish aquaculture in southwestern New Brunswick. Appendix D. pp. 50-54. In: Department of Fisheries and Oceans, Science Branch, Maritimes Region. 2003. Salmon Holding Capacity in Southwestern New Brunswick. Can. Tech. Rep. Fish. Aquat. Sci. 2489: iv + 60 p.
- Robinson, S.M.C. 2004. A roundtable discussion of the future of aquaculture for sea urchins. pp. 387-393. In: Lawrence, J.D. (Ed.) *Sea Urchins- Fisheries and Ecology: Proceedings of the International Conference on Sea Urchin Fisheries and Aquaculture*. Puerto Varas, Chile March 25-27, 2003. DEStech Publications Inc./Lancaster, PA, USA.
- Robinson, S.M.C., J.M Lawrence, L. Burrige, K. Haya, J.D Martin, J.D Castell and A Lawrence. 2004. The effectiveness of different pigment sources in colouring the gonads of the green sea urchin (*Strongylocentrotus droebachiensis*). pp. 215-221. In: Lawrence, J.D. (Ed.) *Sea Urchins- Fisheries and Ecology: Proceedings of the International Conference on Sea Urchin Fisheries and Aquaculture*. Puerto Varas, Chile March 25-27, 2003. DEStech Publications Inc./Lancaster, PA, USA.
- Robinson, S.M.C. 2004. The evolving role of aquaculture in the global production of sea urchins. pp. 343-358. In: Lawrence, J.D. (Ed.) *Sea Urchins- Fisheries and Ecology: Proceedings of the International Conference on Sea Urchin Fisheries and Aquaculture*. Puerto Varas, Chile March 25-27, 2003. DEStech Publications Inc./Lancaster, PA, USA.
- Pearce, C.M., T.L. Daggett and S.M.C. Robinson. 2004. Effect of urchin size and diet on gonad yield and quality in the green sea urchin (*Strongylocentrotus droebachiensis*). *Aquaculture* 233:337-367.
- Auffrey, L.M., S.M.C. Robinson, M.A. Barbeau. 2004. Effect of green macroalgal mats on burial depth of soft-shelled clams (*Mya arenaria* L.). *Mar. Ecol. Prog. Ser.* 278:193-203
- Castell, J.D., E.J. Kennedy, S.M.C. Robinson, G.J. Parsons, T.J. Blair and E. Gonzalez-Duran. 2004. Effect of dietary lipids on fatty acid composition and metabolism in juvenile green sea urchins (*Strongylocentrotus droebachiensis*). *Aquaculture* 242: 417-435
- Daggett, T.L., C.M. Pearce, M. Tingley, S.M.C. Robinson and T. Chopin. 2005. Effect of prepared and macroalgal diets and seed stock source on somatic growth of juvenile green sea urchins (*Strongylocentrotus droebachiensis*). *Aquaculture* 244:263-281.
- Chopin, T.; Robinson, S.; McKinley, R.S.; Fitzgerald, P.; Ugarte, R.; Lander, T.; Banks, J.; Koebberling, U.; Burchill, M. 2004. AquaNet [electronic resource] : integrated aquaculture : an old recycling concept for renewed sustainability = AquaNet : l'aquaculture integree : un concept ancien de recyclage pour renouer avec la durabilite. SH 37 .A68 2004 DVD
- Wong, M.C. M.A. Barbeau, A.W. Hennigar and S.M.C. Robinson. 2005. Protective refuge for seeded juvenile sea scallops (*Placopecten magellanicus*) from sea star (*Asterias* sp.) and crab (*Cancer irroratus*, *Carcinus maenas*) predation: the effect of seeding density and presence of alternative prey. *Can. J. Fish. Aquat. Sci.* 62:1766-1781.
- Lander, T. Barrington, K., Robinson, S., MacDonald, B. and Martin, J. 2004. Dynamics of the blue mussel as an extractive organism in an integrated multi-trophic aquaculture system. *Bull Aquacult. Assoc. Canada* 104-3: 19-28.
- Chopin, T. and S.M.C. Robinson. 2004. Defining the appropriate regulatory and policy framework for the development of integrated multi-trophic aquaculture practices: Introduction to the workshop and positioning of the issues. *Bull Aquacult. Assoc. Canada* 104-3:4-10.
- Chopin, T., Robinson, S.M.C., Sawhney, M., Bastarache, S., Belyea, E., Shea, R., Armstrong, W., Stewart, I., and Fitzgerald, P. 2004. The AquaNet integrated multi-trophic aquaculture project: Rationale of the project and development of kelp cultivation as the inorganic extractive component of the system. *Bull Aquacult. Assoc. Canada* 104-3:11-18.
- Robinson, S.M.C and Chopin, T. 2004. Defining the appropriate regulatory and policy framework for the development of integrated multi-trophic aquaculture practices: Summary of the workshop and issues for the future. *Bull Aquacult. Assoc. Canada* 104-3:73-82.

- Pearce, C.M., S.W. Williams, F.Yuan, J.D. Castell and S.M.C. Robinson. 2005. Effect of temperature on somatic growth and survivorship of early post-settled green sea urchins (*Strongylocentrotus droebachiensis*). *Aquaculture Research* 36:600-609
- Walbourne, C.M., J.F. Piercey, S.M.C. Robinson, J.D. Castell, C.M. Pearce. 2005. Preliminary assessment of recirculation technology for cultivation of the green sea urchin (*Strongylocentrotus droebachiensis*). *Can. Tech. Rep. Fish. Aquat. Sci.* 2569:iv+17p
- Robinson S.M.C., L.M. Auffrey and M.A. Barbeau. 2005. Far-field impacts of eutrophication on the intertidal zone in the Bay of Fundy, Canada with emphasis on the soft-shell clam, *Mya arenaria*. pp.253-274. In: Hargrave, B.T. (Ed.) *The Handbook of Environmental Chemistry Volume 5-M: Water Pollution Environmental Effects of Marine Finfish Aquaculture*. Springer Verlag/Berlin, Heidelberg. 467 p.
- Yeats, P.A., T.G. Milligan, T.F. Sutherland, S.M.C. Robinson, J.A. Smith, P. Lawton, C.D. Levings. 2005. Lithium normalized zinc and copper concentrations in sediments as measures of trace metal enrichment due to salmon aquaculture. Pp. 207-220. In: Hargrave, B.T. (Ed.) *The Handbook of Environmental Chemistry Volume 5-M: Water Pollution Environmental Effects of Marine Finfish Aquaculture*. Springer Verlag/Berlin, Heidelberg. 467 p.
- Kennedy, E.J., S.M.C. Robinson, G.J. Parsons and J.D. Castell. 2005. Effect of protein source and concentration on somatic growth of juvenile green sea urchins (*Strongylocentrotus droebachiensis*) *J. World Aqua. Soc.* 36(3):320-336.
- Lander, T.R., Barrington, K.A., Robinson, S.M.C., MacDonald, B.A., Martin, J.D. 2005. Dynamics of the blue mussel as an extractive organism in an integrated aquaculture system. *Bull. Aquacul. Assoc. Canada*, 104 (3), 19-28.
- Barrington, K., Ridler, N., Chopin, T., Robinson, S., Page, F., MacDonald, B. and Haya, K. 2005. Social Perceptions of Integrated Multi-Trophic Aquaculture. Final report for the Atlantic Canada Opportunities Agency, Saint John, Canada. 80 p.
- Parsons, G.J. and S.M.C. Robinson. 2005. Sea Scallop Aquaculture in the Northwest Atlantic. Chapter 16. pp. 907-944. In: (Shumway, S.E. and G.J. Parsons Eds.) *Scallops: Biology, Ecology and Aquaculture (Revised Edition)*. Elsevier Science B.V., Amsterdam, The Netherlands
- Newell, C. R., C. H. Pilskaln, S. M. Robinson & B. A. MacDonald. 2005. The contribution of marine snow to the particle food supply of the benthic suspension feeder, *Mytilus edulis*. *J. Exp. Mar. Biol. Ecol.* 321:109-124.
- Chopin, T. and S.M.C. Robinson. 2006. Rationale for developing integrated multi-trophic aquaculture (IMTA): an example from Canada. *Fish Farmer Magazine*, Vol. 29, No. 1, p. 20-21, January/ February.
- Robinson, S.M.C., T. Chopin, S. Boyne Travis, J.D. Martin, K. Haya. 2006. A summary of food safety concerns on products grown on an integrated multi-trophic aquaculture (IMTA) site in the Bay of Fundy. Report prepared for the Atlantic Regional Interdepartmental Shellfish Committee (ARISC). November 2006.
- Daggett, T.L., C.M. Pearce and S.M.C. Robinson. 2006. A comparison of three land-based containment systems for use in culturing green sea urchins, *Strongylocentrotus droebachiensis* (Müller) (Echinodermata: Echinoidea). *Aquaculture Res.* 37:339-350
- Ridler, N., Robinson, B., Chopin, T., Robinson, S., and Page, F., 2006. Development of integrated multi-trophic aquaculture in the Bay of Fundy, Canada: a socio-economic case study. *World Aquaculture* 37: 43-48
- Dumont, C.P; Himmelman, J.H; Robinson, S.M.C. 2007. Random movement pattern of the sea urchin *Strongylocentrotus droebachiensis*. *Journal of Experimental Marine Biology and Ecology* 340 (1):80-89.
- Kennedy, E.J., S.M.C. Robinson, G.J. Parsons and J.D. Castell. 2007. Effect of dietary minerals and pigment on somatic growth of juvenile green sea urchins (*Strongylocentrotus droebachiensis*) *J. World. Aqua. Soc.* 38(1):36-48.
- Kennedy, E.J., S.M.C. Robinson, G.J. Parsons and J.D. Castell. 2007. Effect of lipid source and concentration on the somatic growth of juvenile green sea urchins (*Strongylocentrotus droebachiensis*) *J. World. Aqua. Soc.* 38(3):335-352.
- Ridler, N. K. Barrington, B. Robinson, M. Wowchuk, T. Chopin, S. Robinson, F. Page, G. Reid, M. Szemerda, J. Sewuster, S. Boyne-Travis 2007. Integrated Multitrophic Aquaculture -- Canadian Project Combines Salmon, Mussels, Kelps. *Global Aquaculture Advocate* 10(2):52-55.



- Ridler, N., Barrington, K., Wowchuk, M., Chopin, T., Robinson, S., Page, F., Reid, G., Szemerda, M., Sewuster, J., and Boyne-Travis, S., 2007 - Enhancing sustainability of salmon culture in Canada: a socio-economic analysis of multi-trophic aquaculture (IMTA). *Aquaculture Association of Canada Spec. Publ.* 12: 97- 99
- Ridler, N., Wowchuk, M., Robinson, B., Barrington, K., Chopin, T., Robinson, S., Page, F., Reid, G., Szemerda, M., Sewuster, J., and Boyne-Travis, S., 2007 - Integrated multi-trophic aquaculture (IMTA): a potential strategic choice for farmers. *Aquaculture Economics & Management* 11: 99-110.
- Robinson, B., Wowchuk, M., Ridler, N., Chopin, T., Robinson, S., Page, F., Haya, K., and MacDonald, B., 2007 - The socio-economics of integrated multi-trophic aquaculture (IMTA): a case study of the Bay of Fundy. In: *Ocean Management*. N. Catto (Ed.). OMNR, Volume 1 (in press).
- Reid, G.K., Chopin, T., Robinson, S., Neori, A., Buschmann, A.H., Shpigel, M., Rodger, A. and Bolton, J., 2007 - Integrated multi-trophic aquaculture. Wikipedia, The Free Encyclopedia, July 3, 2007. [http://en.wikipedia.org/wiki/Integrated\\_Multi-trophic\\_Aquaculture](http://en.wikipedia.org/wiki/Integrated_Multi-trophic_Aquaculture)
- Chopin, T., S.M.C. Robinson, M. Troell, A. Neori, A.H. Buschmann and J. Fang. 2007. *Ecological Engineering: Multi-Trophic Integration for Sustainable Marine Aquaculture*. In: Jorgensen, S.E. (editor.), *Encyclopedia of Ecology* 3: 2463-2475. Elsevier, Oxford. ISBN 13: 978-0-44-452033-3
- Wildish, D.J., D.D. Kristmanson , S.M.C. Robinson. 2008. Does skimming flow reduce population growth in horse mussels? *Journal of Experimental Marine Biology and Ecology* 358:33–38.
- Chopin, T. and S.M.C. Robinson 2008. Pour une aquaculture durable: un changement s'impose – FrancVert Le Webzine Environnementale Printemps 2008 5(2). 6p. <http://www.francvert.org/pages/52dossierpouruneaquaculturedurable.asp>
- González-Durán, E., J.D. Castell, S.M.C. Robinson and T.J. Blair. 2008. Effects of dietary lipids on the fatty acid composition and lipid metabolism of the green sea urchin *Strongylocentrotus droebachiensis*. *Aquaculture* 276:120-129
- Chopin, T., Robinson, S., Barrington, K., Reid, G., Ridler, N., Robinson, B., Wowchuk, M., Sawhney, M., Page, F., Haya, K., Burrige, L., Szemerda, M., Sewuster, J., and Boyne-Travis, S., 2008 - Integrated multi-trophic aquaculture – Acuicultura multi-trófica integrada. *Panorama Acuicola Magazine* 13 (5): 22-29.
- Reid, G.K., M. Liutkus, S.M.C. Robinson, T.R. Chopin, T. Blair, T. Lander, J. Mullen, F. Page, R.D. Moccia. 2009. A review of the biophysical properties of salmonid faeces: implications for aquaculture waste dispersal models and integrated multi-trophic aquaculture. *Aqua. Res.* 40:257-273
- MacDonald, B.A., S.M.C. Robinson and K.A. Barrington, K.A 2009. Potential use of exhalent siphon area inn estimating feeding activity of blue mussels, *Mytilus edulis*. *J. Shellfish Res.* 28:1-9.
- Shuve, H., E. Caines, N. Ridler, T. Chopin, G. K. Reid, M. Sawhney, J. Lamontagne, M. Szemerda, R. Marvin, F. Powell, S. Robinson, S. Boyne-Travis 2009. Survey Finds Consumers Support Integrated Multitrophic Aquaculture - Effective Marketing Concept Key. *Global Aquaculture Advocate* March/April:12-13
- Barrington, K., N. Ridler, T. Chopin, S.M.C. Robinson, B. Robinson. 2010. Social aspects of the sustainability of integrated multi-trophic aquaculture. *Aquaculture International* 18 (2):201-211.
- Lander, T.R., R.K. Shaw, S.M.C., Robinson and J.D. Martin. 2010. Blue mussel (*Mytilus edulis*) settlement patterns on antifoulant treated salmon nets and its implications for recycling used salmon nets for mussel spat collection in Integrated Multi-trophic Aquaculture (IMTA). *Can. Tech. Rep. Fish. Aquat. Sci.* 2848: vi + 20 p.
- Robinson, S.M.C., J.D. Castell, C.M. Walbourne and C.M. Pearce. 2010 The importance of beta-carotene for somatic growth of juvenile green sea urchins (*Strongylocentrotus droebachiensis*) pp. 397-402. In: (Harris, L.H., S.A. Bottger, C.W. Walker & M.P. Lesser eds) *Echinoderms: Durham. Proceedings of the 12th International Echinoderm Conference, 7-11 August 2006, Durham, New Hampshire, U.S.A.* 679p. CRC Press, Taylor & Francis Group, New York
- Pearce, C. and S.M.C. Robinson. 2010. Recent Advances in Sea-Urchin Aquaculture and Enhancement in Canada. *Bull. Aquacul. Assoc. Canada* 108-1: 38-48.
- Reid, G.K., M. Liutkus, A. Bennett, S.M.C. Robinson, B. MacDonald, F. Page. 2010. Absorption efficiency of blue mussels (*Mytilus edulis* and *M. trossulus*) feeding on Atlantic salmon (*Salmo salar*) feed and fecal particulates: Implications for integrated multi-trophic aquaculture. *Aquaculture* 299:165-169.

- Daggett, T.L., C.M. Pearce, S.M.C. Robinson, T. Chopin. 2010. Does method of kelp (*Saccharina latissima*) storage affect its food value for promoting somatic growth of juvenile green sea urchins (*Strongylocentrotus droebachiensis*)? *J. Shellfish Res.* 29(1):247-252.
- MacDonald, B.A. S.M.C. Robinson, K.A. Barrington 2011. Feeding activity of mussels (*Mytilus edulis*) held in the field at an Integrated Multi Trophic Aquaculture (IMTA) site (*Salmo salar*) and exposed to fish food in the laboratory. *Aquaculture* (in press).
- Robinson, S.M.C., J.D. Martin, J.A. Cooper, T.R. Lander, G.K. Reid, F. Powell and R. Griffin. 2011. The role of three dimensional habitats in the establishment of integrated multi-trophic aquaculture (IMTA) systems. *Bull. Aquacul. Assoc. Canada* 109-2: 23-29.
- Hamer, A., Martin, J.L., Robinson, S., Page, F., Hill, B., Powell, F. and Justason, A. 2012. Spatial and temporal trends in paralytic shellfish poisoning levels in the soft shell clam, *Mya arenaria*, along the southwestern coast of New Brunswick in the Bay of Fundy. *Can. Tech. Rep. Fish. Aquat. Sci.* 2982: viii +97p
- Browdy, C.L., Hulata, G., Liu, Z., Allan, G.L., Sommerville, C., Passos de Andrade, T., Pereira, R., Yarish, C., Shpigel, M., Chopin, T., Robinson, S., Avnimelech, Y. & Lovatelli, A. 2012. Novel and emerging technologies: can they contribute to improving aquaculture sustainability? In R.P. Subasinghe, J.R. Arthur, D.M. Bartley, S.S. De Silva, M. Halwart, N. Hishamunda, C.V. Mohan & P. Sorgeloos, eds. *Farming the Waters for People and Food. Proceedings of the Global Conference on Aquaculture 2010, Phuket, Thailand. 22–25 September 2010.* pp. 149–191. FAO, Rome and NACA, Bangkok
- Liutkus, M., S. Robinson, B. MacDonald and G. Reid. 2012. Quantifying the effects of diet and mussel size on the biophysical properties of the blue mussel, *Mytilus* spp., feces egested under simulated IMTA conditions. *J. Shellfish Res.*, 31(1): 1–9.
- Lander, T.R., S.M.C. Robinson, B.A. MacDonald, and J.D. Martin. 2012. Enhanced growth rates and condition index of blue mussels (*Mytilus edulis*) held at integrated multi-trophic aquaculture (IMTA) sites in the Bay of Fundy. *J. Shellfish Res.* 31: 997-1007.
- Nelson, E.J., MacDonald, B.M., Robinson, S.M.C. 2012 A Review of the Northern Sea Cucumber *Cucumaria frondosa* (Gunnerus, 1767) as a Potential Aquaculture Species. *Reviews in Fisheries Science* 20:212-219.
- Nelson, E.J., MacDonald, B.M., Robinson, S.M.C. 2012. The absorption efficiency of the suspension-feeding sea cucumber, *Cucumaria frondosa*, and its potential as an extractive integrated multi-trophic aquaculture (IMTA) species. *Aquaculture* 370-371:19-25.
- Graydon, C.M., S.M.C. Robinson, J.A. Cooper and R.E. Scheibling. 2012. Detection of canthaxanthin in the tissues of *Mytilus edulis* and *Strongylocentrotus droebachiensis* as a potential tracer to determine the zone of influence of salmon aquaculture. *Aquaculture* 366–367:90–97
- Chopin, T., MacDonald, B., Robinson, S., Cross, S., Pearce, C., Knowler, D., Noce, A., Reid, G., Sawhney, M., Ang, K.P., and Backman, C., 2012 - The Canadian Integrated Multi-Trophic Aquaculture Network (CIMTAN) – A network for a new ERA of Ecosystem Responsible Aquaculture. *Fisheries* (in press).
- Martínez-Espiñeira, R., Chopin, T., Robinson, S., Noce, A., Yip, W., and Knowler, D., 2012 - Estimation of the biomitigation benefits of Integrated Multi-Trophic Aquaculture. Technical report to the Atlantic Canada Opportunities Agency – Atlantic Innovation Fund (ACOA-AIF), 212 p.
- Saksida, S., Bricknell, I., Robinson, S. and Jones, S. 2012. Sea lice monitoring and non-chemical measures: C. Population ecology and epidemiology of sea lice in Canadian waters. *Canadian Science Advisory Secretariat Report:* 38p.
- St. Hilaire, S., Robinson S.M.C., Glebe, B. and Cox R. 2012. Sea lice monitoring and non-chemical measures: F. Non-chemical measures of control and prevention. *Canadian Science Advisory Secretariat Report:* 15p.
- Robinson, S.M.C. and Reid, G.R. 2012. A review of the potential near-field and far-field effects of integrated multi-trophic aquaculture (IMTA) practices in the Bay of Fundy with emphasis on the organic extractive trophic level (ETL) component. *Canadian Science Advisory Secretariat Report:* 67p.

- Bartsch A., Robinson S.M., Liutkus M., Ang K.P., Webb J., Pearce C.M. 2013. Filtration of sea louse, *Lepeophtheirus salmonis*, copepodids by the blue mussel, *Mytilus edulis*, and the Atlantic sea scallop, *Placopecten magellanicus*, under different flow, light and copepodid-density regimes. *J. Fish Dis.* 36(3):361-370.
- Lander, T.R., S.M.C. Robinson, B.A. MacDonald, and J.D. Martin. 2013. Characterization of the suspended organic particles released from salmon farms and their potential as a food supply for the suspension feeder, *Mytilus edulis* in Integrated Multi-trophic Aquaculture (IMTA) systems. *Aquaculture* 406-407, 160-171.
- Webb J.L., Vandenbor, J., Pirie, B., Robinson, S.M.C., Cross, S.F., Jones, S.R.M. and Pearce, C.M.. 2013. Effects of temperature, diet, and bivalve size on the ingestion of sea lice (*Lepeophtheirus salmonis*) larvae by various filter-feeding shellfish. *Aquaculture* 406-407: 9–17
- Irisarri, J., Fernandez-Reiriz, M. J., Robinson, S. M. C., Cranford, P. J., and Labarta, U. 2013. Absorption efficiency of mussels *Mytilus edulis* and *Mytilus galloprovincialis* cultured under Integrated Multi-Trophic Aquaculture conditions in the Bay of Fundy (Canada) and Ria Ares-Betanzos (Spain). *Aquaculture* 388: 182-192.
- Reid, G.K., T. Chopin, S.M.C. Robinson, P. Azevedo, M. Quinton, E. Belyea. 2013. Weight ratios of the kelps, *Alaria esculenta* and *Saccharina latissima*, required to sequester dissolved inorganic nutrients and supply oxygen for Atlantic salmon, *Salmo salar*, in Integrated Multi-Trophic Aquaculture systems. *Aquaculture* 408-409: 34-46.
- Reid, G.K., S.M.C. Robinson, T. Chopin and B.A. MacDonald. 2013. Dietary proportion threshold of fish culture solids required by organic extractive species to reduce the net organic load in open-water Integrated Multi-Trophic Aquaculture (IMTA) systems: A scoping exercise with co-cultured Atlantic salmon (*Salmo salar*) and blue mussel (*Mytilus edulis*). *J. Shellfish Res.* (in press)
- Chopin C., MacDonald, B., Robinson, S., Cross, S., Pearce, C., Knowler, D., Noce, A., Reid, G., Cooper, A., Speare, D., BurrIDGE, L., Crawford, C., Sawhney, M., Ang, K.P., Backman, C., Hutchinson, M. 2013. The Canadian Integrated Multi-Trophic Aquaculture Network (CIMTAN)—A Network for a New Era of Ecosystem Responsible Aquaculture. *Fisheries* 38:297-308.
- Cranford, P.J., Reid, G.K. and Robinson, S.M.C. 2013. Open water integrated multi-trophic aquaculture: constraints on the effectiveness of mussels as an organic extractive component. *Aqua Env Inter.* 4: 163-173.
- Cranford, P.J., Duarte P., Robinson, S.M.C., Fernández-Reiriz, M.J., Labarta, U. 2014. Suspended particulate matter depletion and flow modification inside mussel (*Mytilus galloprovincialis*) culture rafts in the Ría de Betanzos, Spain. *Journal of Experimental Marine Biology and Ecology* 452: 70–81.
- Robinson, S.M.C., and Reid, G.K. 2014. Review of the potential near-and far-field effects of the organic extractive component of Integrated Multi-Trophic Aquaculture (IMTA) in Southwest New Brunswick with emphasis on the blue mussel (*Mytilus edulis*). DFO Can. Sci. Advis. Sec. Res. Doc. 2014/026. vii + 60 p.
- Filgueira, R., C. Byron, L.A. Comeau, B. Costa-Pierce, P.J. Cranford, J. Grant, T. Guyondet, H.M. Jansen, C.W. McKinsey, J.K. Petersen, G.K. Reid, S.M.C. Robinson, A. Smaal, R. Sonier, Ø. Strand, T. Strohmeier. 2015. An integrated ecosystem approach for assessing the potential role of bivalve shells as part of the carbon trading system. *Marine Ecology Progress Series* 518: 281–287
- Irisarri J., Fernández-Reiriz, M.J., Labarta U., Cranford P.J., Robinson, S.M.C. 2015. Availability and utilization of waste fish feed by mussels *Mytilus edulis* in a commercial Integrated Multi-Trophic Aquaculture (IMTA) system: a multi-indicator assessment approach. *Ecological Indicators* 48:673–686.
- Brager L.M., Cranford P.J., Grant J, Robinson S.M.C. 2015. Spatial distribution of suspended particulate wastes at open-water Atlantic salmon and sablefish aquaculture farms in Canada. *Aquaculture Environment Interactions* 6: 135-49.
- Martínez-Espiñeira R, Chopin T, Robinson S, Noce A, Knowler D, Yip W. 2015. Estimating the biomitigation benefits of Integrated Multi-Trophic Aquaculture: A contingent behavior analysis. *Aquaculture* 437: 182-194.
- Cubillo A.M., Ferreira J.G., Robinson S.M.C., Pearce C.M., Corner R.A., Johansen J. 2016. Role of deposit feeders in integrated multi-trophic aquaculture — A model analysis. *Aquaculture*. 453: 54-66.
- Jansen, H.M., Reid, G.K., Bannister, R.J., Husa, V., Robinson, S.M.C., Cooper, J.A., Quinton, C. & Strand, O. 2016. Discrete water quality sampling at open-water aquaculture sites: limitations and strategies. *Aquaculture Environment Interactions*, 8: 463-480.

- Wildish, D.J. & Robinson, S.M.C. 2016. A new secondary ecotope for talitrids: driftwood in the Bay of Fundy. *Crustaceana*, 89: 737-757.
- Wildish, D.J. & Robinson, S.M.C. 2016. Ultimate cause(s) of dwarfism in invertebrates: the case of driftwood talitrids. *Evolutionary Ecology Research*, 17: 685-698.
- Yu, Z.H., Robinson, S.M.C., Xia, J.J., Sun, H.Y. & Hu, C.Q. 2016. Growth, bioaccumulation and fodder potentials of the seaweed *Sargassum hemiphyllum* grown in oyster and fish farms of South China. *Aquaculture*, 464: 459-468.
- Nelson, E.J., Robinson, S.M.C., Feindel, N., Sterling, A., Byrne, A. & Pee Ang, K. 2017. Horizontal and vertical distribution of sea lice larvae (*Lepeophtheirus salmonis*) in and around salmon farms in the Bay of Fundy, Canada. *Journal of Fish Diseases*.
- Byrne, A.A., Pearce, C.M., Cross, S.F., Jones, S.R.M., Robinson, S.M.C., Hutchinson, M.J., Miller, M.R., Haddad, C.A. & Johnson, D.L. 2018. Planktonic and parasitic stages of sea lice (*Lepeophtheirus salmonis* and *Caligus clemensi*) at a commercial Atlantic salmon (*Salmo salar*) farm in British Columbia, Canada. *Aquaculture*, 486, 130-138.
- Byrne, A.A., Pearce, C.M., Cross, S.F., Jones, S.R.M., Robinson, S.M.C., Hutchinson, M.J., Miller, M.R., Haddad, C.A. & Johnson, D.L. 2018. Field assessment of Pacific oyster (*Crassostrea gigas*) growth and ingestion of planktonic salmon louse (*Lepeophtheirus salmonis*) larvae at an Atlantic salmon (*Salmo salar*) farm in British Columbia, Canada. *Aquaculture*, 490, 53-63.
- Wildish, D.J., Pavesi, L. & Robinson, S.M.C. 2018. Comparing oxygen uptake rates of driftwood and wrack generalist talitrid amphipods. *Marine and Freshwater Behaviour and Physiology*, 51, 203-211.
- Baillie, S.M., McGowan, C., May-McNally, S., Leggatt, R., Sutherland, B.J.G., and Robinson, S. 2019. Environmental DNA and its applications to Fisheries and Oceans Canada: National needs and priorities. *Can. Tech. Rep. Fish. Aquat. Sci.* 3329: xiv + 84 p
- Reid, G.K., Lefebvre, S., Filgueira, R., Robinson, S.M.C., Broch, O.J., Dumas, A., and Chopin, T.B.R. 2020. Performance measures and models for open-water integrated multi-trophic aquaculture. *Reviews in Aquaculture* 12(1): 47-75.
- Sardenne, F., Simard, M., Robinson, S.M.C., and McKindsey, C.W. 2020. Consumption of organic wastes from coastal salmon aquaculture by wild decapods. *Science of the Total Environment* 711.
- Yu, Z., Robinson, S., MacDonald, B., Lander, T., and Smith, C. 2020. Effect of diets on the feeding behavior and physiological properties of suspension-feeding sea cucumber *Cucumaria frondosa*. *Journal of Oceanology and Limnology* 38(3): 883-893.
- Frühe, L., V. Dully, D. Forster, N. B. Keeley, O. Laroche, X. Pochon, S. Robinson, T. A. Wilding and T. Stoeck 2021. Global trends of benthic bacterial diversity and community composition along organic enrichment gradients of salmon farms. *Frontiers in Microbiology* 12(853). 10.3389/fmicb.2021.637811
- Wildish, D. J., S. M. C. Robinson and M. Black 2021. Locomotor activity rhythms of North Atlantic coastal talitroids. *Marine and Freshwater Behaviour and Physiology*, 1-22. 10.1080/10236244.2021.1993737
- MacGregor, KA., M.F. Lavoie, S.M.C. Robinson, É. Simard, and C.W. McKindsey. 2023 Lab and field evaluation of tagging methods for the use of acoustic telemetry to observe sea urchin movement behaviour at ecologically relevant spatio-temporal scales. *Animal Biotelemetry* 11(1): 3.